was treated with 35 g. (0.343 mole) of acetic anhydride, added portionwise while vigorously shaking and cooling to keep the temperature below 40°. Precipitation was completed by adding 50 ml. of 6N hydrochloric acid. The solids were removed by filtration, dried and taken up in 500 ml. of hot methanol. After filtering off inorganic salts and cooling, 5.8 g. of yellow needles were obtained, m.p. 216– 217.5°; concentration of their filtrate to 50 ml. yielded an additional 1.9 g. Recrystallization of these combined crops from methanol gave 6.7 g. (75%) of small yellow needles, m.p. 218–219°.

Anal. Caled. for $C_{11}H_{10}O_3S$: C, 59.44; H, 4.53; S, 14.43. Found: C, 59.68; H, 4.46; S, 14.16.

p-Thiomyristoxycinnamic acid. p-Thioleinnamic acid, 11.8 g. (0.066 mole) and 1 g. of potassium hydroxide were dissolved in 200 ml. of dry pyridine and the reaction mixture was stirred for 1 hr. Myristoyl chloride, 22.7 g. (0.092 mole) was added dropwise during 1 hr. below 30°, and stirring was continued for 3 additional hr. The solids obtained upon dilution with 500 ml. of water and 100 ml. of 6N hydrochloric acid were removed by filtration and then shaken with 200 ml. methanol to remove excess myristic acid. The residue was recrystallized from benzene with the aid of charcoal, 6.8 g. (27%) light yellow plates, m.p. 166-167°.

Anal. Caled. for C22H34O3S: C, 70.70; H, 8.77; S, 8.21. Found: C, 70.55; H, 8.88; S, 8.00.

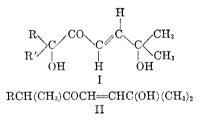
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Structure of the Side Chain of Cucurbitacin B

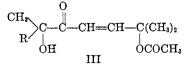
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In two recent Communications,^{1,2} Lavie and his co-workers have proposed the partial structure I



for elatericin A and α -elaterin. Structure II was proposed by Rivett and Enslin³ for elaterin and for cucurbitacins A and C. Still more recently⁴ partial structure III has been proposed for elaterin



D. Lavie and Y. Shvo, Proc. Chem. Soc., 220 (1958).
D. Lavie, Y. Shvo, and D. Willner, Chem. and Ind., 1361 (1958).

and cucurbitacins A, B, and C. From our work we also have concluded that curcurbitacin B (formerly called fabacein II⁵) has structure III, and we wish to report results which confirm this structure and which clarify the results of catalytic hydrogenation.

Cucurbitacin B has been assigned the molecular formula $C_{32}H_{48}O_8^6$ and is a monoacetate, containing an α,β -unsaturated carbonyl group.^{5,7} Analyses of an apparently identical product isolated from the juice of *Echinocystis fabacea* were reported^{5,8} to agree with the formulas $C_{30}H_{44-46}O_8$ (Calcd. for $C_{30}H_{44}O_8$: C, 67.64; H, 8.33; for $C_{30}H_{46}O_8$: C, 67.39; H, 8.67. Found: C, 67.38; H, 8.40; average of 12 analyses). However the formulas $C_{32}H_{46-48}O_8 \cdot 0.5$ H₂O (Calcd.: C, 67.70; H, 8.34; or C, 67.46; H, 8.67) also are equally satisfactory and agree better with the analyses of derived products.

When cucurbitacin B was hydrogenated in 95% ethanol using 10% palladium on carbon as catalyst, from 1.3 to 1.6 moles of hydrogen was absorbed.⁸ The resulting solution contained acetic acid, and titration indicated that 0.3 to 0.6 mole of acetic acid was formed. Evidently hydrogenolysis as well as hydrogenation had occurred. The paper chromatogram of the hydrogenated material showed that two products were present which could be separated readily. One product is dihydrocucurbitacin B (VI) (Caled. for C₃₂H₄₈O₈: C, 68.54; H, 8.63; for C₃₂H₅₀O₈: C, 68.30; H, 8.96. Found: C, 68.38; H, 8.80); m.p. 163-164° from acetonehexane; $[\alpha]_{D}^{25} + 57^{\circ}$ (c = 0.91 in CHCl₃); UV (ethanol) λ_{max} 282 m μ , log ϵ 2.32; IR (CHCl₃): 2.92 (OH), 5.79 w(AcO), 5.85 sh (C=O), 5.89 (C=O), 8.10 (AcO). The other product is dihydrodeacetoxycucurbitacin B (VII) (Calcd. for $C_{30}H_{46}O_6$: C, 71.68; H, 9.22; for C₃₀H₄₈O₆: C, 71.39; H, 9.59. Found: C, 71.59; H, 9.37); m.p. 208-210° from ether; $[\alpha]_{D}^{25} + 57^{\circ}$ (c = 0.93 in CHCl₃); UV (ethanol) λ_{max} 279 m μ , log ϵ 2.46; IR (CHCl₃): 2.92 (OH), 5.85 sh (C=O), 5.90 (C=O). In both products the α,β -unsaturated carbonyl system present in cucurbitacin B^{5,7} has disappeared.

When cucurbitacin B in acetic acid was treated wi h zinc dust for 4 hr. at room temperature, a new product was obtained. The ultraviolet and infrared spectra showed that the α,β -unsaturated carbonyl system had disappeared. Although cucurbitacin B gave only a pale yellow color with tetranitromethane, the new compound gave an orange color, indicating the production of a highly alkylated isolated double bond. Analysis showed that the new

⁽³⁾ D. E. A. Rivett and P. R. Enslin, *Proc. Chem. Soc.*, 1958, in press. We wish to thank Dr. Enslin for sending us a copy of this Communication.

⁽⁴⁾ Private communication from Dr. P. R. Enslin summarizing a note sent to Chemistry and Industry in December, 1958.

⁽⁵⁾ W. O. Eisenhut and C. R. Noller, Abstracts of paper presented at the San Francisco meeting of the American Chemical Society, April 1958.

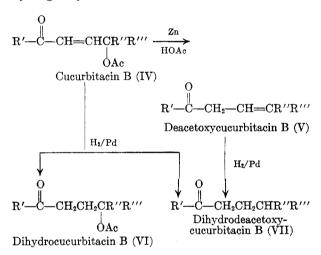
⁽⁶⁾ D. E. A. Rivett and F. H. Herbstein, Chemistry and Industry, 393 (1957).

⁽⁷⁾ P. R. Enslin, S. Rehm, and D. E. A. Rivett, J. Sci. Food Agr., 8, 673 (1957).

⁽⁸⁾ W. O. Eisenhut and C. R. Noller, J. Org. Chem., 23, 1984 (1958).

compound was a deacetoxycucurbitacin B (V) (Caled. for $C_{30}H_{44}O_6$: C, 71.97; H, 8.86; for $C_{30}H_{46}O_6$: C, 71.68; H, 9.22. Found: C, 71.72; H, 9.05); m.p. 178–179°; $[\alpha]_D^{25} + 78^\circ$ (c = 0.76 in ethanol); UV (ethanol) λ_{max} 292 m μ , log ϵ 2.66; IR (CHCl₃) 2.91 (OH), 5.87 (C=O), 5.90 (C=O). Thus, by this reaction not only has the acetoxy group been removed, but the double bond no longer is in conjugation with the carbonyl group. Hydrogenation of this product converted it quantitatively into a single compound which was identical in all respects with dihydrodeacetoxycucurbitacin B (VII).

These results can be explained by partial formula IV for cucurbitacin B. The easy loss of acetic acid from curcurbitacin B by mild saponification⁵ or hydrogenolysis indicates that either an enol acetate



or allyl acetate grouping is present. The former is excluded by the lack of absorption around 5.68 μ , which is typical of enol acetates. It should be noted that hydrogenation of cucurbitacin B stops completely when considerably less than two moles of hydrogen is absorbed. Thus hydrogenolysis of some molecules takes place previous to hydrogenation of the double bond, and hydrogenation of the double bond without hydrogenolysis occurs simultaneously (VII and VI). Once the double bond is hydrogenated, hydrogenolysis no longer occurs. Facile deacylation of cucurbitacin by reduction with zinc and migration of the double bond out of conjugation with the carbonyl group to give deacetoxycucurbitacin B (V) is analogous to the easy removal by zinc reduction of a hydroxyl or acetoxyl group α to a carbonyl group and undoubtedly occurs by a similar mechanism.⁷

Oxidation of dihydrodeacetoxycucurbitacin B (VII) in aqueous dioxane with sodium periodate gave about equal amounts of neutral and acidic products. The latter were converted to the methyl esters with diazomethane and steam distilled. Gas chromatography using a Carbowax-impregnated

column identified the chief component of the volatile esters as methyl isocaproate. Methyl isocaproate was obtained also when alkaline hydrogen peroxide was used as an oxidizing agent. Thus the carbon skeleton of isocaproic acid is present as a side chain. The presence of a 1,2-diketo or α hydroxy keto grouping at the sixth and seventh carbon atoms of the side chain also is indicated.

Acetvlation of cucurbitacin B and of its hvdrogenation products with acetic anhydride in pyridine gave only amorphous compounds which appeared to be homogeneous on paper. They were purified for analysis by selecting a narrow cut when passed through a chromatographic column. Cucurbitacin B gave a tetraacetate (VIII) (Calcd. for $C_{38}H_{52}O_{11}$: C, 66.65; H, 7.65; O, 25.70; for $C_{38}H_{54}O_{11}$: C, 66.45; H, 7.92; O, 25.63. Found: C, 66.42; H, 7.83; O, 25.38); m.p. 125–130°; $[\alpha]_{D}^{25}$ +4.4 (c = 0.95 in CHCl₃); UV (ethanol) λ_{max} 228 m μ , log ϵ 4.33; IR (CHCl₃) 2.90 (OH), 5.80 (AcO), 5.92 (C=O), 6.13 (C=C), 8.10 (AcO). Thus cucurbitacin B contains three acetylatable hydroxyl groups and at least one nonacetylatable hydroxyl group which probably is tertiary.

Dihydroeucurbitacin B also gave a tetraacetate (IX) (Calcd. for $C_{38}H_{54}O_{11}$: C, 66.45; H, 7.92; for $C_{38}H_{56}O_{11}$: C, 66.26; H, 8.19. Found: C, 66.52; H, 8.16); m.p. 120–125°; $[\alpha]_D^{25}$ –16.3 (c = 0.92 in CHCl₃); UV (ethanol) λ_{max} 286 m μ , log ϵ 2.28; IR (CHCl₃) 2.90 (OH), 5.80 (AcO), 5.90 sh (C=O), 8.10 (AcO). As expected, dihydrodeacetoxycucurbitacin B (VII) gave a triacetate (X). (Calcd. for $C_{36}H_{52}O_9$: C, 68.76; H, 8.34; for $C_{36}H_{54}O_9$: C, 68.54; H, 8.63. Found: C, 68.55; H, 8.46); m.p. ~130°; $[\alpha]_D^{25}$ –20.4° (c = 1.37 in CHCl₃); UV (ethanol) λ_{max} 282 m μ , log ϵ 2.44; IR (CHCl₃) 2.90 (OH), 5.80 (AcO).

Oxidation of all three acetates with chromium trioxide in acetic acid at room temperature gave a mixture of volatile acids and a large neutral fraction. From 0.5 to 1.5 moles of carbon dioxide also was evolved. Examination of the volatile acids by paper chromatography and of their methyl esters by gas chromatography showed that the acetate of dihydrodeacetoxycucurbitacin B (X) gave chiefly isocaproic acid. The acetate of cucurbitacin B (VIII), however, gave chiefly β , β -dimethylacrylic acid. These facts together with the observations concerning the hydrogenation products of cucurbitacin B, prove that its acetoxy group must be in the side chain and that R'' and R''' are methyl groups. The neutral products of the oxidations appear to be methyl ketones (positive iodoform reaction), which, coupled with the results of periodate oxidation, indicates that R' in formula IV CH3

may be replaced by the group $R \xrightarrow{\downarrow C} H$ leading OH

to the partial formula III for cucurbitacin B. The formation of $\beta_{,\beta}$ -dimethylacrylic acid would

⁽⁷⁾ Cf. R. B. Woodward, F. Sondheimer, D. Taub, K. Heusler, and W. M. McLamore, J. Am. Chem. Soc., 74, 4225 (1952).

The neutral fractions from the oxidations of all of the acetylated compounds appear to be identical and to consist of a mixture of at least six substances. The component present in largest quantity, which for the present is designated as ketone A, appears to have the formula C₂₈H₃₈O₈ (Calcd.: C, 66.91; H, 7.62. Found: C, 67.10; H, 7.47); m.p. 219-221° from acetone-hexane; $[\alpha]_{\rm D}^{25}$ +86.6 (c = 0.96 in CHCl₃); UV (ethanol): λ_{max} 243 m μ , log ϵ 4.1; 335 m μ , log ϵ 2.0; IR (CHCl₃) 5.77 (AcO), 5.85 (C=O), 6.00, 6.15 (C=C-C=O), 8.12 (AcO). Analyses of the component present in next largest amount, designated as ketone B, indicate the formula C₂₈H₃₆O₈ (Calcd.: C, 67.18; H, 7.25; Found: C, 67.09; H, 7.15); m.p. 154–157° from acetone hexane; $[\alpha]_{D}^{25}$ -84.9 (c = 1.14 in CHCl₃); UV (ethanol) λ_{max} 295 mµ, log ϵ 2.37; IR (CHCl₃) 5.78 (AcO), 5.91 (C=O), 8.10 (AcO). Both compounds gave a positive iodoform reaction, indicating the possible presence of methyl ketone groups.

The remaining four neutral products, which are present in smaller amounts, have been isolated in a fairly pure state but have not yet been further **,**

investigated. Inasmuch as they do not appear when the oxidations are carried out for longer periods, it is possible that they are intermediates in the formation of ketones A and B.

Enslin and Rivett report the isolation of two methyl ketones designated as cucurbitones A and C from the chromic acid oxidation products of the diacetates of cucurbitacins A and C.³ Their analyses and molecular weight determination on cucurbitone A, m.p. 210°, $[\alpha]_D + 100°$ (CHCl₃), λ_{max} 245 m μ (log ϵ 4.04), correspond to the molecular formula C₃₀H₃₈O₁₀. Their analyses of cucurbitone C, m.p. 246-247°, $[\alpha]_D + 153$ (CHCl₃) λ_{max} 241 (log ϵ 4.1), support the molecular formula C₃₀H₄₀O₉. The physical constants of our methyl ketone A and cucurbitone A are similar, but the analyses are very different. The other important product of our oxidations, ketone B, is certainly different than any previously reported.

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